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## Short communication

# Reversed-phase C<sub>18</sub> high-performance liquid chromatography of gibberellins GA<sub>3</sub> and GA<sub>1</sub><sup>-1</sup>

Grolamys Castillo\*, Servando Martinez

Cuban Research Institute of Sugar Cane By-products (ICIDCA), Via Blanca 804 Carretera Central, San Miguel del Padrón, P.O. Box 4026, Ciudad Habana, C.P. 11000, Cuba

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## **Abstract**

Gibberellins (GAs) are a group of plant growth hormones that are involved in the regulation of shoot elongation in many crop plants and the regulation of flowering in some biennials or photoperiodic plants. A reversed-phase HPLC procedure was applied for the separation of GA<sub>3</sub> and GA<sub>1</sub> with high resolution by means of ionic suppression, without the necessity for derivatization, using UV detection in fermentation broths. The reproducibility at these chromatographic conditions showed a 2% R.S.D. for GA<sub>3</sub>, the recovery of GA<sub>3</sub> was higher than 95% and the detection limit for GA<sub>3</sub> standard solution was ca. 50 ng. GA<sub>3</sub> and GA<sub>1</sub> concentrations were determined for different fermentation cultures of the fungus Gibberella fujikuroi. © 1997 Elsevier Science B.V.

Keywords: Gibberellins; Plant hormones

#### 1. Introduction

Gibberellins (GAs) belong to an important group of plant growth hormones involved in the control of different physiological processes such as: regulation of shoot elongation in many crop plants, regulation of flowering in some biennials or photoperiodic plants, fruit set, fruit external ripening and fruit development, and others [1–3].

At present, plant growth hormones are applied in agriculture to improve crop quality and yields. Industrial production of these plant growth hormones have been possible using fermentative processes from the action of some fungus on specific substrates

and different methods have been established for their recovery and purification from fermentation medium [4].

High-performance liquid chromatography (HPLC) is now a routine procedure for the purification and separation of gibberellins (GAs). Reversed-phase C<sub>18</sub> HPLC of gibberellins has been the most frequently used and reported HPLC system for separating free GAs [1-3,5-10]. The HPLC of some gibberellin derivatives have also been reported [6]. Different chromatographic methods have been developed for the determination of GAs such as: thin-layer chromatography (TLC) [11], gas chromatography (GC) [11], gas chromatography—mass spectrometry (GC–MS) [1,3,9] and gas chromatography—selected ion monitoring MS (GC–SIM-MS) [1,2].

According to the method developed by Barendse et al. in [7] we modified it for quantitative de-

<sup>\*</sup>Corresponding author.

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termination of  $GA_3$  and  $GA_1$  in a fermentation culture of the fungus *Gibberella fujikuroi*. An isocratic reversed-phase  $C_{18}$  HPLC of  $GA_3$  and  $GA_1$  is described in this paper. The ionic suppression technique, which involves the regulation of pH, was considered suitable for separating GAs, without the necessity for derivatization, using UV detection.

# 2. Experimental

# 2.1. High-performance liquid chromatography

The HPLC system consisted of the following components from Philips analytical (Cambridge, UK): a pump (PU 4100) and a variable-wavelength UV-Vis detector (PU 4110). The injector was a Rheodyne (Cotati, CA, USA) Model 7125. The column was a reversed-phase C<sub>18</sub> column Spherisorb S5ODS1 (25 cm×4.6 mm I.D., 5 µm, Philips Scientific, Cambridge, UK). The mobile phase consisted of 30% methanol (HPLC; Panreac Montplet y Esteban, Barcelona, Spain), containing 0.01 M H<sub>2</sub>PO<sub>4</sub>, adjusted with KOH to pH 3. The detection took place at 206 nm. Routine sample calculations were based on the comparison of peak areas with external standard peak areas using BIOCROM software for chromatography (CIGB, Havana, Cuba). The chromatographic conditions are given in the figure legends.

## 2.2. Gibberellins

GA<sub>3</sub> was purchased from Sigma (St. Louis, MO, USA) and GA<sub>1</sub> was provided by the ICIDCA's Biochemical Laboratory. GA<sub>3</sub> and GA<sub>1</sub> standard solutions were prepared by dissolving them in the mobile phase. Calibration curves were developed for a GA<sub>3</sub> concentration range from 0.1 to 1 mg/ml and GA<sub>1</sub> concentration range from 0.01 to 0.5 mg/ml.

## 2.3. Sample preparation

Cultures were filtered through Whatman No 1 paper. The filtrates were adjusted to pH 2.5 and diluted when necessary in mobile phase so that the detector response was within the concentration range used to prepare the calibration curve. The filtrates

were then filtered by ISO DISC Syringe Tip Filter Unit 0.45  $\mu m$ . The sample was injected in 20  $\mu l$  portions.

## 3. Results and discussion

A reversed-phase HPLC procedure was developed for the separation of Gibberellins  $GA_3$  and  $GA_1$ . The linearity of the analysis was evaluated for  $GA_3$  and  $GA_1$  calibration curve concentration range. A slope of 248.78 with a standard error of 0.65 and the origin as intercept were obtained for  $GA_3$  standard curve for n=5. A slope of 104.73 with an standard error of 0.97 and the origin as intercept were obtained for  $GA_1$  standard curve for n=5. The correlation coefficients calculated for the compound concentration versus the peak area were 0.9999 (P < 0.05) for  $GA_3$  and 0.9996 (P < 0.05) for  $GA_1$ . The detection limit of this HPLC technique was ca. 50 ng on column for  $GA_3$  standard solution.

A 1.49 resolution was achieved with Spherisorb S5ODS1 column but it was not necessary working at the lower limit of pH (2.3) as in the method described by Barendse. This corroborates the statement by Barendse et al. in Ref. [7] that by choosing a particular methanol concentration, containing 0.01 M H<sub>3</sub>PO<sub>4</sub> and an appropriate pH, a particular set of GAs can be analyzed with good resolution.

The more polar compounds elute sooner from reversed-phase  $C_{18}$  HPLC [5,6]. The gibberellin having a double bond (GA<sub>3</sub>) is more polar than the saturated analogue. Fig. 1 shows the separation of standards of GA<sub>3</sub> and GA<sub>1</sub>. The retention times were 11.00 min for GA<sub>3</sub> and 12.62 min for GA<sub>1</sub>.

A good analytical within-day precision was obtained for GA<sub>3</sub> in a fermentation culture. The results obtained were  $\bar{X}$  (mg GA<sub>3</sub>/ml)=0.381,  $\sigma_{n-1}$ =7×  $10^{-3}$  and R.S.D.=2.2% for n=5.

To evaluate the recovery of  $GA_3$ , a fermentation culture of known  $GA_3$  concentration was taken and  $GA_3$  standard was added at two different concentrations: 0.1 and 0.5 mg/ml. The sample was submitted to the described sample preparation procedure and recoveries of 96.4% and 98.7% were found, respectively.

Fig. 2 shows a typical chromatogram of a fermentation culture. At this wavelength no signifi-

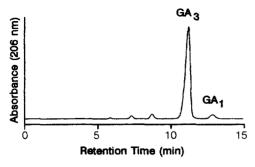


Fig. 1. Chromatogram of  $C_{18}$  HPLC of  $GA_1$  and  $GA_3$  (20  $\mu$ l, 1  $\mu$ g/ $\mu$ l). Column: Spherisorb S5ODS1, 25 cm×4.6 mm I.D. Mobile phase: 30% MeOH, containing 0.01 M H<sub>3</sub>PO<sub>4</sub> adjusted with KOH to pH 3; flow-rate 1 ml/min. Detection at 206 nm, 1.0 a.u.f s.

cant interferences for GA<sub>3</sub> and GA<sub>1</sub> from contaminants in the acidic fermentation culture filtrate have been observed. This agrees with the results obtained by J.-T. Lin and E. Heftmann [5].

Results from different fermentation cultures achieved with the method described in this paper are presented in Table 1.

The results shown in this table are obtained under different fermentation conditions such as: pH, fermentation technique, scaling-up, and others, used in the optimization study of the fermentative process conditions.

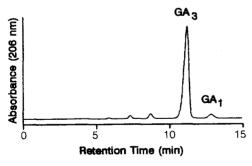


Fig. 2. Chromatogram of C<sub>18</sub> HPLC of a fermentation culture (20 μl). Column: Spherisorb S5ODS1, 25 cm×4.6 mm I.D. Mobile phase: 30% MeOH, containing 0.01 M H<sub>3</sub>PO<sub>4</sub> adjusted with KOH to pH 3; flow-rate 1 ml/min. Detection at 206 nm, 1.0 a.u.f.s.

Table 1  $GA_3$  and  $GA_1$  concentrations from different fermentation cultures

Sample	Concentrations (mg/ml)	
	$\overline{GA_3}$	GA <sub>1</sub>
1	0.425	0.099
2	0.301	0.063
3	0.388	0.171
4	0.229	0.029
5	0.415	0.058

For HPLC conditions see Fig. 2.

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## References

- [1] T.I. Potter, K.P. Zanewich, S.B. Rood, Plant Growth Regul. 12 (1993) 133.
- [2] J.-T. Lin, A.E. Stafford, G.L. Steffens, Agric. Biol. Chem. 55 (1991) 2183.
- [3] G.L. Steffens, J.-T. Lin, A.E. Stafford, J.D. Metzger, J.P. Hazebroek, J. Plant Growth Regul. 11 (1992) 165.
- [4] P.K.R. Kumar, B.K. Lonsane, Advances in Applied Microbiology 34 (1989) 29.
- [5] J.-T. Lin, E. Heftmann, J. Chromatogr. 213 (1981) 507.
- [6] J.-T. Lin, A.E. Stafford, J. Chromatogr. 452 (1988) 519.
- [7] G.W.M. Barendse, P.H. van de Werken, N. Takahashi, J. Chromatogr. 198 (1980) 449.
- [8] J.-T. Lin, A.E. Stafford, J. Chromatogr. 543 (1991) 471.
- [9] S.W. Johnson, R.C. Coolbaugh, Plant Physiol. 94 (1990) 1696.
- [10] M.L. Brenner, Ann. Rev. Plant Physiol. 32 (1981) 511.
- [11] R.Ch. Rachev, S.V. Bojkova, R. Pavlova-Rouseva, V.K. Gancheva, M.P. Dimitrova, J. Chromatogr. 437 (1988) 287.